Aridlands Ecology Laboratory
2010.06.04, S.Castle

Chlorophyll-*a* Double-Extraction with Methanol

Materials

Mortar and pestle
15mL screw-cap vials
Polystyrene cuvettes, Fisher # 14-377-010
5000µL and 1000 µL pipettes and pipette tips
Spectrophotometric grade Methanol
Ammonium Acetate (solid form)
Plastic funnel
Orbital shaker

Solutions:

1. Ammonium Acetate Solution:
2. *Aqueous Methanol Solution:* Add 20mL of Ammonium Acetate Solution to a 1000mL flask. Fill to the line with 100% Methanol cover with parafilm or a blue cap and invert to mix.

Processing and Extraction:

Note: All processing should be done in minimal light to help prevent degradation of chlorophylls.

1. Grind soil sample in mortar and pestle until homogenous, and place 3g soil into labeled 15mL screw-cap vial.
2. Add 9mL of Methanol solution to each vial and shake gently by hand.
3. Place all vials horizontally on shaker and let them shake for 5 hours, samples should be covered and protected from light.
4. After shaking, centrifuge samples at 3000rpm for 6 minutes.
5. Carefully pour supernatant into a separate labeled vial. This sample is ready for analysis on the spectrophotometer.

*If samples or extracts need be held overnight, they must be kept in the fridge at 4°C.

6. Repeat steps 2-5 for second extraction.

Determining Chlorophyll-*a* Concentration on the Spectrophotometer:

Warm up the Spec:

1. Plug in the Spec.
2. The spectrophotometer takes approximately 10-15minutes to warm up. During this time it will run through a self test.
3. Turn on the lamp. Press the "Vis" button until you see "Vis" appear in the digital screen.
4. When the spec is ready the screen will read a wavelength (λ) of 486. Align the cuvette tray using a new cuvette with a sticky note rolled up inside of it (green sticky notes work best). If the door is propped slightly open you can see the blue light hitting the cuvette holder. The holder should be positioned so that the light hits the center of the cuvette.

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Calibrating the Spec:

1. Calibration should be done using the same solution that the samples were extracted with. This means extra solution should be made during the sample extraction process.

Cuvettes should be handled with care. Dirty cuvettes will give bad readings. Only touch the foggy side of the cuvette.

2. Place a cuvette with blank solution into the cuvette holder. Close door.
3. Type in the wavelength push the "λ" button and then push "calibrate". Calibrate for all wavelengths that you will be measuring at. Calibration can be done for up to 10 wavelengths. The absorbance reading should be zero for all calibrated points.

Measuring Samples:

1. When measuring samples the door must be completely closed and the cuvette holder aligned.
2. Only use new and clean cuvettes for running samples. They are not that expensive and are not worth cleaning.
3. Make sure that the extraction solution is compatible with the cuvette material. Acetone etches polystyrene, so certain precautions should be taken when using acetone.
4. Using the 5000µL pipette, transfer 3mL of extract to cuvette.
5. Promptly measure the absorbance at 652, 665nm and 750nm.

When finished with readings turn Spec off by unplugging the instrument.

Calculating Chla concentrations:

1. Use the following equation to determine concentration of chlorophyll-*a* in each soil sample:

$$\mu g\ \text{Chl-}a\ g^{-1} = \frac{(-8.9062 * A652) - (16.5169 * A665) * V}{L * G}$$

where:

A652 = initial absorbance = (abs. at 652nm) – (abs. 750nm)
A665 = initial absorbance = (abs. at 665nm) – (abs. 750nm)
V = volume of extract (milliliters)
L = light path length in cuvette (centimeters)
G = amount of soil (grams)

Calculation Reference: Ritchie, 2006